



## Spiroazetidines in Drug Discovery

### Key Points

- Offer more interactions with target proteins
- Provide superior physiochemical properties
- Offer novel intellectual properties

### Overview

Spiroazetidines, like all spirocyclic scaffolds, are bestowed with three advantages over their flat counterparts: inherent three-dimensional structures may offer more interactions with target proteins; spiroazetidines may provide superior physiochemical properties thus more drug-like; and novel structures may offer fresh intellectual properties. Meanwhile, although no spiroazetidine-containing drugs are currently approved for marketing, at least two of them have been advanced to clinical trials. Their applications in medicinal chemistry are destined to grow, especially since many of them are now commercially available.

**PharmaBlock**

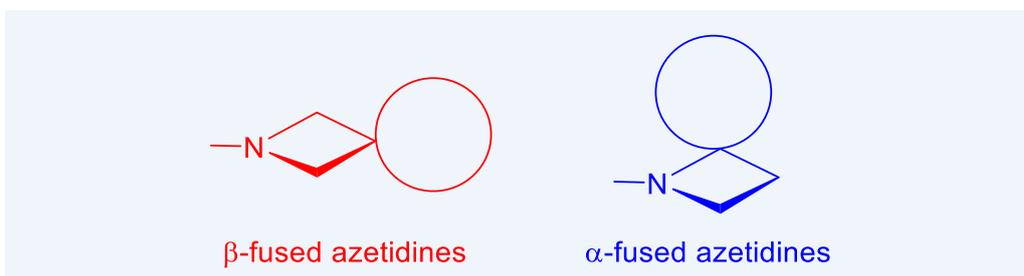
PharmaBlock designs and synthesizes over 4038 Spiros and 775 Spiros products are in stock. [CLICK HERE](#) to find detailed product information on webpage.



Spirocycles are making more and more frequent appearances in drugs as both core structures and peripheries of drug molecules. They have at least three advantages as drug fragments:

- Spirocyclic scaffolds are inherently three-dimensional and can project functionalities in all three dimensions to interact more extensively with the protein target of interest and lower off-target effects in comparison to their two-dimensional counterparts;
- They are  $sp^3$ -carbon-rich with high value of *fraction of saturated carbon* ( $F_{sp^3}$ ) and  $sp^3$ -carbon-rich molecules are correlated with favorable physicochemical properties such as higher aqueous solubility;<sup>1</sup> and
- Spirocycles often offer new chemical space to create novel intellectual properties, as amply documented by Zheng et al.<sup>2</sup>

With regard to bicyclic spiroazetidines, the other ring may be fused to azetidine at either the  $\alpha$ - or  $\beta$ -position of the nitrogen atom. Overwhelming examples in the literature are the  $\beta$ -fused azetidine spirocycles.



### Spiroazetidine-containing Drugs

Only one spiroazetidine-containing drug is on the market in Japan: delgocitinib (Corectim, **1**). Discovered by Japanese Tobacco, it is a pan-JAK inhibitor approved as a topical treatment of atopic dermatitis.

Janus kinases (JAKs) recruit signal transducers and activators of transcription (STATs) to cytokine receptors, leading to modulation of gene expression. They are intracellular tyrosine kinases that mediate the signaling of numerous cytokines and growth factors involved in the regulation of immunity, inflammation, and hematopoiesis. There are four members of the Janus kinase family: JAK1, JAK2, JAK3, and TYK2.

## PharmaBlock Products



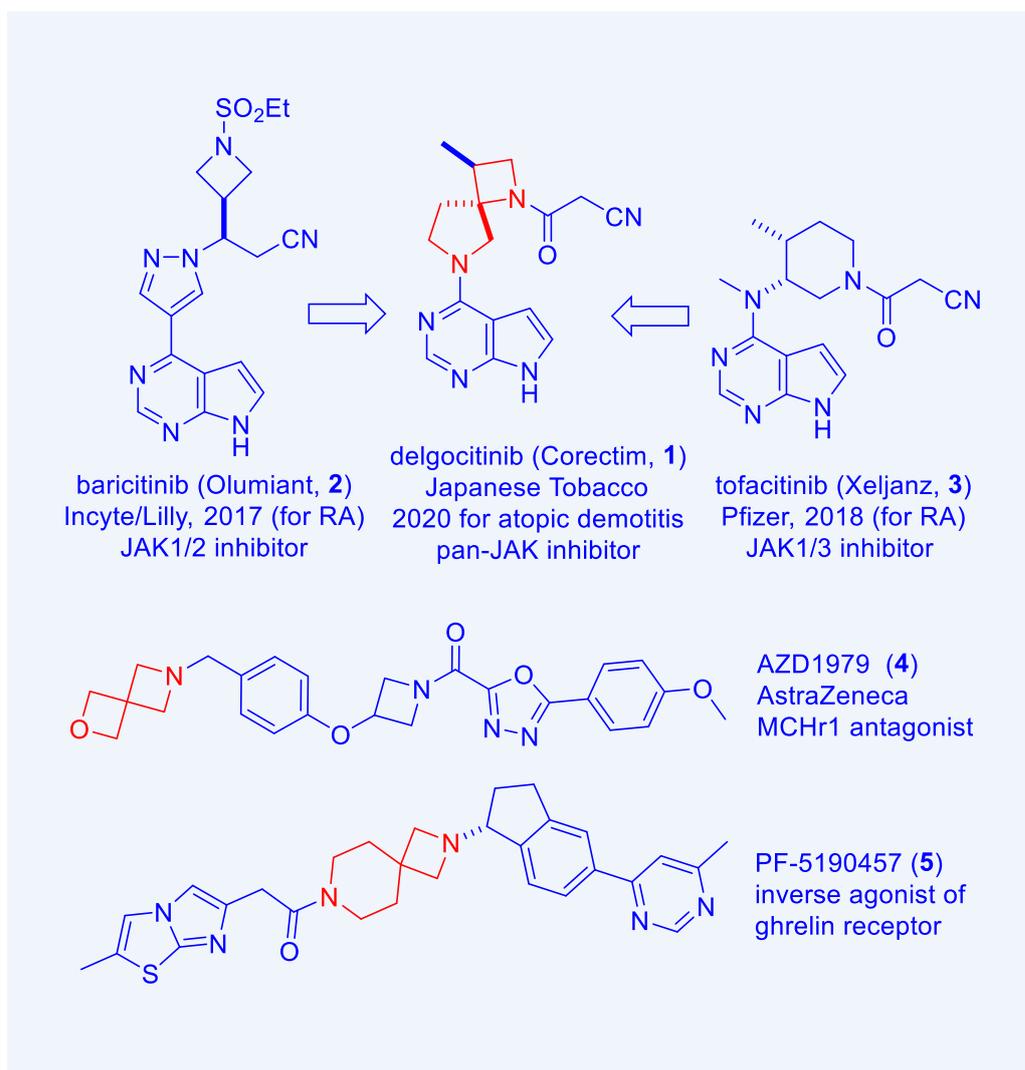
PBZ0617



PBZS2138

Incyte's JAK1/2 dual inhibitor baricitinib (Olumiant, **2**) was approved in 2017 for treating rheumatoid arthritis (RA). Also marketed as a treatment of RA since 2012, Pfizer's tofacitinib (Xeljanz, **3**) is a JAK1/JAK3 inhibitor with moderate activity on JAK2.

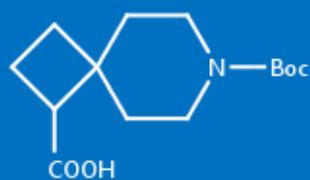
Drug discovery is extremely hard. Yet it can be comparatively easy if you know what you are doing, especially in the field of "me-too" drugs, also known as "patent busting" or euphorically "scaffold hopping" tactics. Pharmaceutical industry flourished on the tail of "me-too" beta blockers. Japanese Tobacco wisely infused features from both baricitinib (**2**) and tofacitinib (**3**) and arrived at delgocitinib (**1**) with a spirocyclic azetidine-pyrrolidine (1,6-diazaspiro[3.4]-octane) fragment.<sup>3</sup> The spiroazetidine moiety closely mimics the three-dimensional head pieces of its progenies: baricitinib (**2**) and tofacitinib (**3**). The discovery of delgocitinib (**1**) showcased the third merit for spirocycles: creating novel IPs.



## PharmaBlock Products

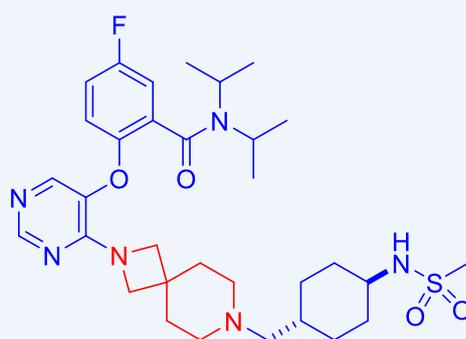


PB00717



PBZ3122

At least three spiroazetidines-containing drugs have been advanced to clinical trials. One is AstraZeneca's melanin concentrating hormone receptor 1 (MCHR1) antagonist AZD1979 (**4**) for the treatment of diabetes.<sup>4</sup> Its peripheral azetidine-oxetane (2-oxa-6-azaspiro[3.3]-heptane) spirocycle is an isostere for morpholine. Another spiroazetidines-containing drug in clinical trials is Pfizer's inverse agonist of the ghrelin receptor (GR) PF-5190457 (**5**) for treating obesity.<sup>5</sup> It has a spirocyclic azetidine-piperidine backbone. The third spiroazetidines-containing drug in clinical trials is Vitae's VTP50469 (**6**) for treating leukemia.<sup>6</sup> It is an inhibitor of the menin and MLL interaction. It also has a spirocyclic azetidine-piperidine framework. Its three-dimensional structure is a good solution for the challenge of tackling protein-protein interactions.



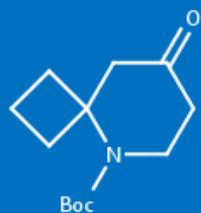
VTP50469 (**6**)  
Inhibitor of  
menin/ MLL  
interaction

## Spiroazetidines in Drug Discovery

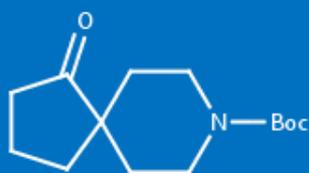
### a. Spiroazetidines as bioisosteres

Spiroazetidines, like all spirocyclic scaffolds, are inherently three dimensional and offer structural novelty. In terms of isosterism, 2,6-diazaspiro[3.3]heptane can mimic piperazine and 2-oxa-6-azaspiro[3.3]heptane bears striking resemblance to morpholine. In the same vein, 2-azaspiro[3.3]heptane has served as an excellent isostere of piperidine and has found several important applications in drug discovery. Therefore, these spiroazetidines have found utility in drug discovery as isosteres for piperazine, morpholine, and piperidine, respectively.

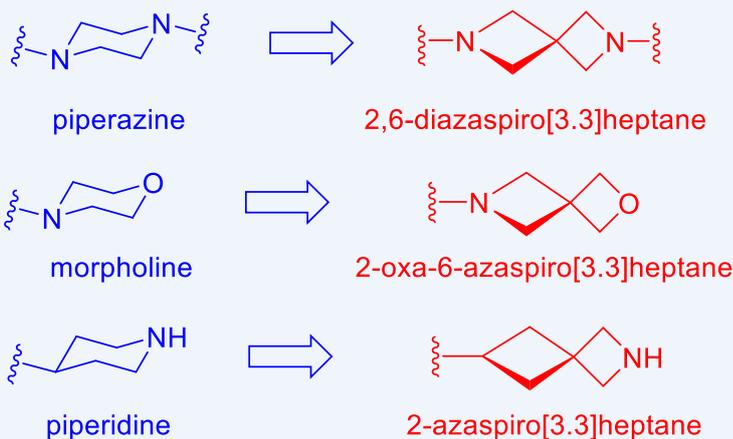
## PharmaBlock Products



PBN20120736



PB05069



Inhibition of ketohexokinase (KHK, also known as fructokinase) promotes fructose metabolism; thus, KHK inhibitors have potential as diabetes treatment. From high-throughput screening (HTS) and further optimization guided by structure-based drug design (SBDD), Maryanoff and coworkers arrived at pyrimidinopyrimidine piperazine **7** as a potent, selective human hepatic KHK (KHK-C isoform) inhibitor. Since piperazine is considered a conformationally constrained isostere for ethyldiamine, 2-oxa-6-azaspiro[3.3]heptane is even further constrained than piperazine. Along this line of speculation, spiroazetidine analog **8** was prepared and tested to have similar enzymatic potency in inhibiting recombinant human hepatic KHK-C as piperazine **7**, indicating that conformational rigidity is tolerated for the structure-activity relationship (SAR) for this pyrimidinopyrimidine series of KHK-C selective inhibitors. Furthermore, both piperazine **7** and spiroazetidine **8** exhibited reasonably potent cellular KHK inhibition (400 and 360 nM, respectively), which relates to their intrinsic potency versus KHK and their ability to enter cells.<sup>7</sup>

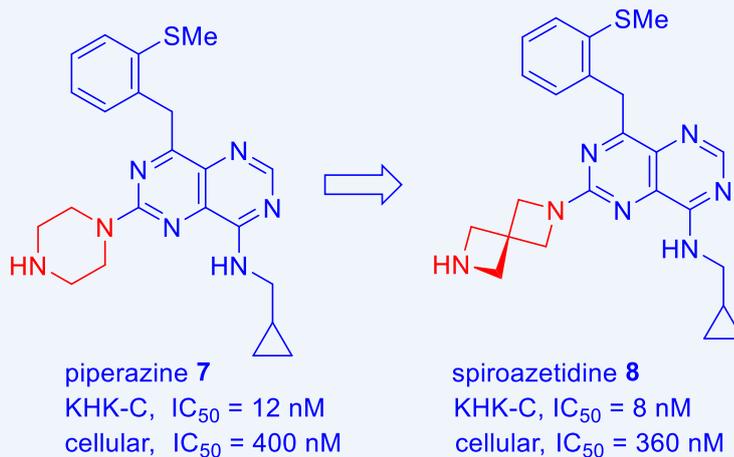
## PharmaBlock Products



PBN20121195



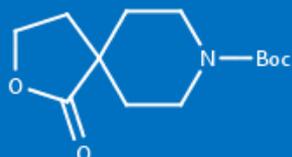
PB06178



Like KHK inhibitors, stearoyl-CoA desaturase (SCD) inhibitors are also potential treatments of metabolic disorders such as diabetes and obesity. To minimize adverse events associated with eyes and skin, liver-targeted SCD inhibitors are preferred. Merck Frost arrived at piperazine **9** as a liver-selective SCD inhibitor with an enzymatic potency of 28 nM against rat SCD. The strategic presence of polar acid moiety was important because carboxylic acids and tetrazoles are recognized organic anionic transporter proteins (OATPs), thus providing the desired *in vivo* properties, i.e., a high liver concentration (target organ for efficacy) and a low systemic concentration to minimize exposures in off-target tissues and cell-associated adverse events (eyes and skin).<sup>8</sup>

Spiroazetidine isostere **10** has a similar enzymatic potency as its prototype **9**. Moreover, both of them are also virtually inactive in a whole cell assay in a human hepatocellular carcinoma line (HepG2), indicating that both spiroazetidine **10** and piperazine **9** do not enter off-target cells via passive diffusion since HepG2 cells are devoid of active OATPs. Gratifyingly, both spiroazetidine **10** and piperazine **9** do cross cell membranes through active transporters when assessed in a rat hepocyte (Rat Hep, which contains functional OATPs) assay.<sup>8</sup>

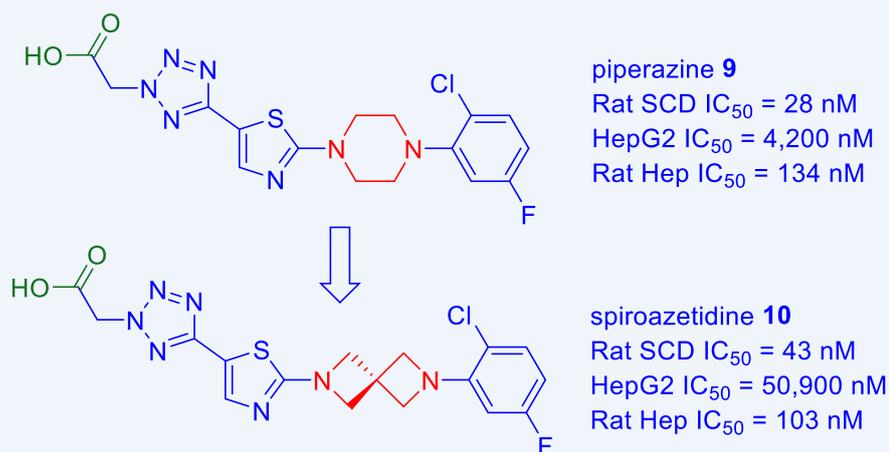
## PharmaBlock Products



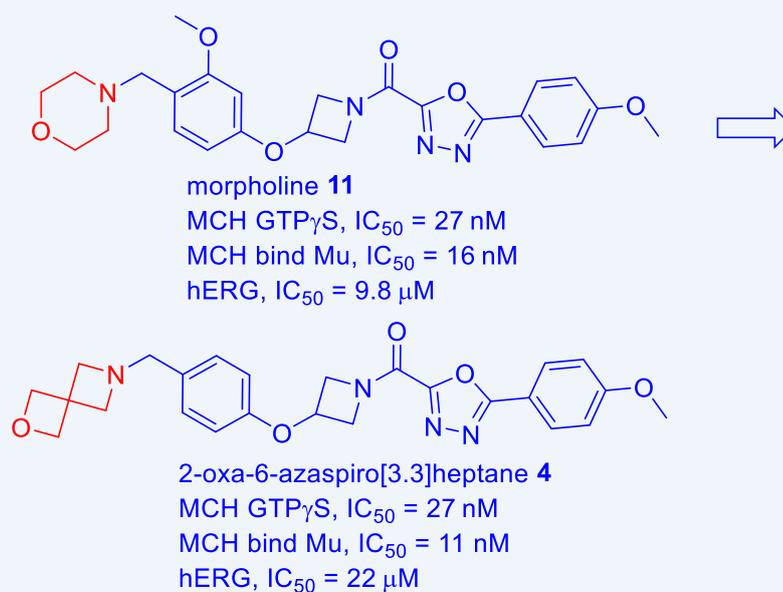
PB07465



PBWBD0221



Continuing the theme of diabetes treatments, MCHR1 antagonists have been explored for weight control. AstraZeneca discovered a series of oxadiazole-containing compounds as represented by morpholine **11**. Their efforts culminated in clinical candidate AZD1979 (**4**) with a novel peripheral spiroazetidine moiety. Spiroazetidine **4** displayed appropriate lipophilicity for a CNS indication, showed excellent permeability with no efflux, and possessed good off-target selectivity, including hERG. Preclinical good-laboratory practice (GLP) toxicology and safety pharmacology studies were without findings and AZD1979 (**4**) was taken into clinical trials.<sup>4</sup>



## PharmaBlock Products

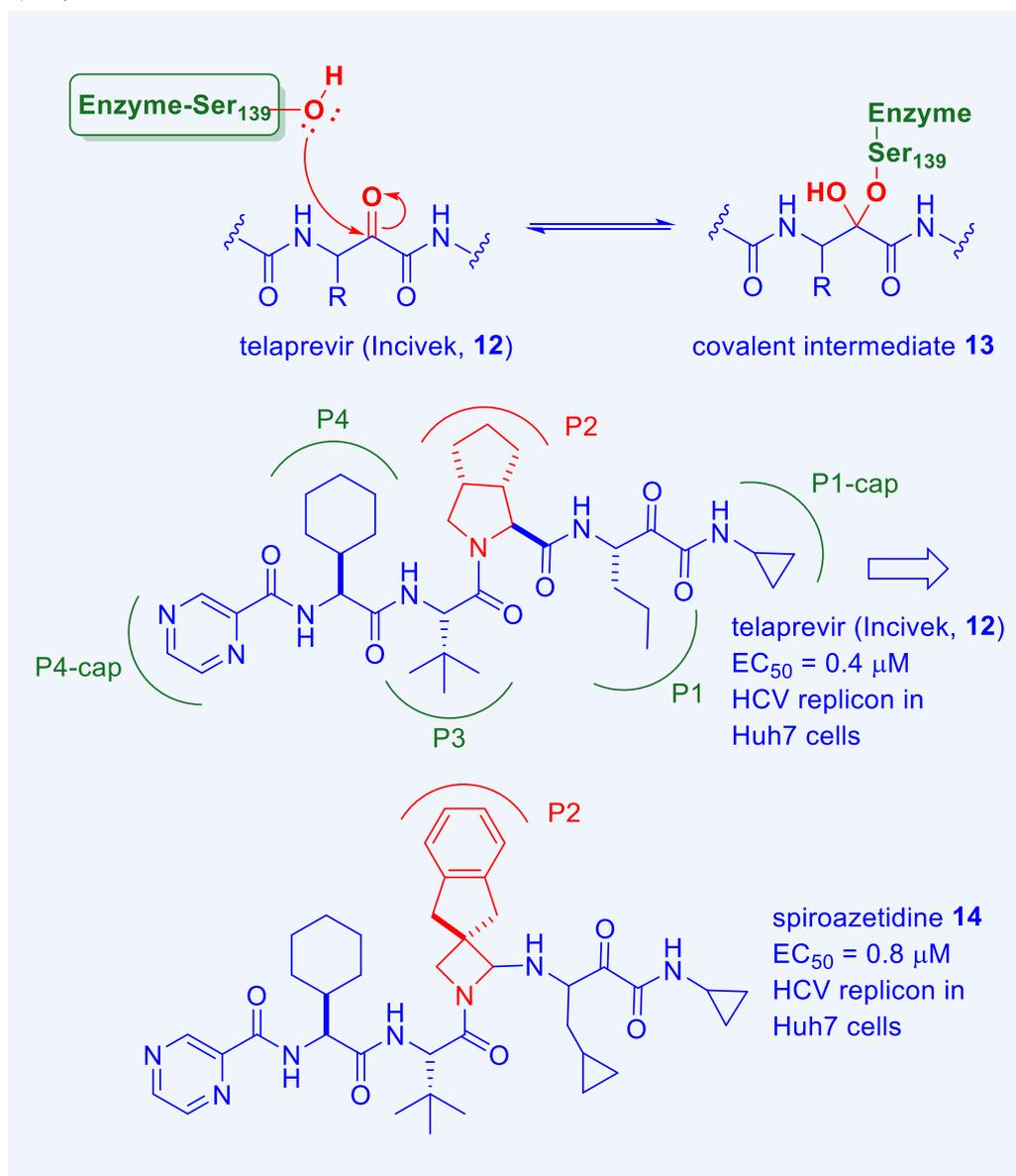


PBN20122077



PB08298

Similar to Schering–Plough’s boceprevir (Victrelis), Vertex’s telaprevir (Incivek, **12**) is a hepatitis C virus (HCV) NS3/4A *serine* protease inhibitor. They are also competitive, covalent reversible inhibitors. Working in concert with His<sub>57</sub> and Asp<sub>81</sub>, the Ser<sub>139</sub> on HCV NS3/4A serine protease enzyme adds to the ketone warhead on telaprevir (**12**) to form a covalent tetrahedral hemiacetal intermediate **13**, which closely mimics the transition state of the hydrolysis processes by the serine protease.<sup>9</sup> Introduction of spiroazetidine moieties at the P2 unit of telaprevir (**12**) resulted in inhibitors with good potency as measured using inhibition of HCV RNA replication in Huh7 cells in a subgenomic HCV replicon system. In particular, spiroazetidine **14** displayed a potency (EC<sub>50</sub> = 0.8 μM) similar to that of telaprevir (**12**, EC<sub>50</sub> = 0.4 μM).<sup>10</sup>



## PharmaBlock Products



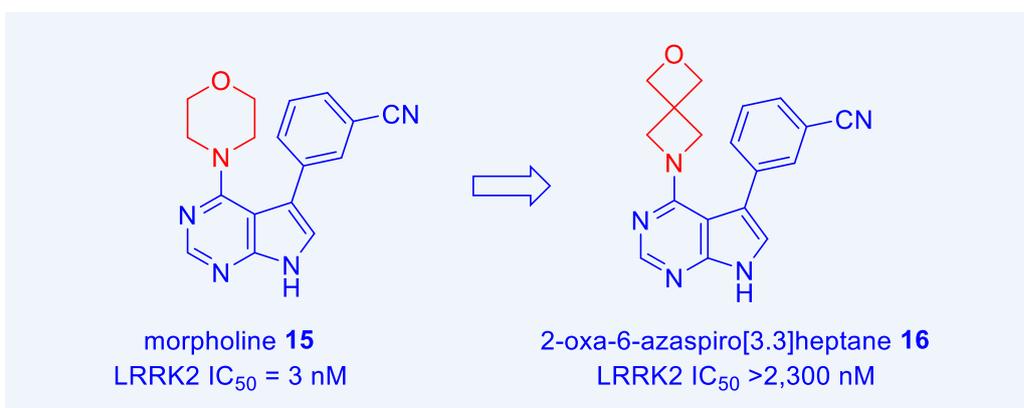
PB97357-1



PB97358-1

Not surprisingly, not all spiroazetidines worked as a panacea.

Leucine rich repeat kinase 2 (LRRK2) inhibitors are potentially useful in the treatment of Parkinson's disease (PD). From HTS and lead optimization, Pfizer arrived at morpholine **15** (PF-06447475) as a highly potent (LRRK2 enzymatic assay:  $IC_{50} = 3 \text{ nM}$ ), selective, brain penetrant, and *in vivo* active LRRK2 inhibitor. Attempt to replace the morpholino group with the 2-oxa-6-azaspiro[3.3]heptane isostere led to spiroazetidine **16**, which was virtually inactive toward LRRK2, regrettably.<sup>11</sup>



### b. Utility of spiroazetidines in medicinal chemistry

Spirocyclic azetidine–piperidine (2,7-diazaspiro[3,5]nonane) fragment served as the backbone of a series of inverse agonists of the ghrelin receptor (GR) discovered by Pfizer.<sup>12,13,5</sup>

GR is a G-protein coupled receptor (GPCR) that plays a role in obesity and glucose homeostasis. Starting from an HTS hit, azetidine–piperidine **17** was discovered as a potent GR *inverse agonist* (hGR  $IC_{50} = 4.6 \text{ nM}$ ,  $K_i = 7.0 \text{ nM}$ , an *inverse agonist* is a drug that binds to the same receptor as an agonist but induces a pharmacological response opposite to that of the agonist) with 43% bioavailability in Sprague–Dawley rat. But it suffered from an undesired off-target effect; namely it displayed muscarinic acetylcholine receptor (mAChR) M2 activity ( $K_i = 269 \text{ nM}$ ).<sup>12</sup>

Conformational restriction of the right-hand portion of compound **17** led to chiral indane **18**, which maintained its potency (hGR  $pK_i = 8.2$ , i.e.,  $K_i = 6.3 \text{ nM}$ ) with improved selectivity over mAChR M2 ( $pK_i = 4.85$ , i.e.,  $K_i = 14.1 \text{ }\mu\text{M}$ ) as well as high receptor occupancy.<sup>13</sup>

## PharmaBlock Products

Absolute



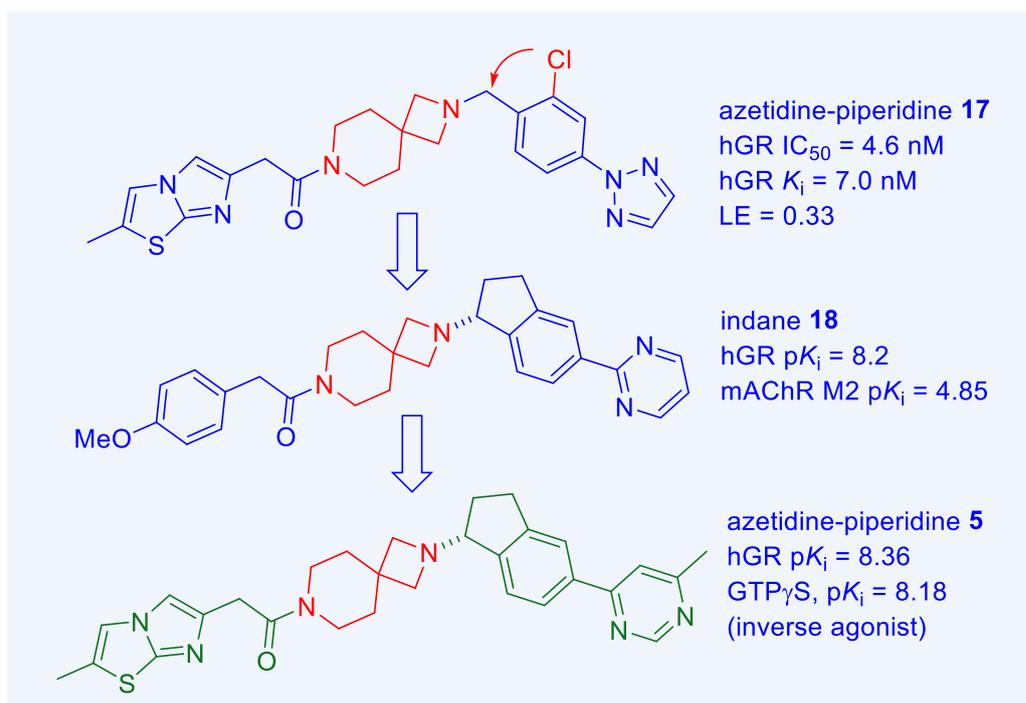
PBWB100120

Absolute



PBWB100431-1

Eventually, meticulous fine-tuning of the SAR culminated in PF-5190457 (**5**,  $pK_i = 8.36$ , i.e.,  $K_i = 4.4$  nM), which showed a better balance of receptor activity and off-target selectivity (M2  $K_b$ /GR  $K_i$  ratio = 266). As a potent, selective, and orally bioavailable GR inverse agonist, azetidine–piperidine **5** was advanced to clinical trials for the treatment of diabetes on the basis of its promising pharmacological and safety profile.<sup>5</sup>



The azetidine–piperidine (2,7-diazaspiro[3,5]nonane) fragment also found success in the fatty acid amide hydrolase (FAAH) inhibitors program as well. FAAH is an integral membrane serine hydrolase responsible for the degradation of fatty acid amide signaling molecules such as endocannabinoid anandamide (AEA), which has been shown to possess cannabinoid-like analgesic properties. Therefore, FAAH inhibitors are explored as treatment of pain. A group of chemists at Janssen prepared heteroarylurea FAAH inhibitors with dozens of diamine linkers. One of them, 2,7-diazaspiro[3,5]nonane **19**, was found to be a potent FAAH inhibitor (hFAAH  $IC_{50} = 8$  nM). In addition, it was found to inhibit FAAH centrally, elevate the brain levels of three fatty acid ethanolamides [FAAs: AEA, oleoyl ethanolamide (OEA) and palmitoyl ethanolamide (PEA)], and was moderately efficacious in a rat model of neuropathic pain.<sup>14</sup>

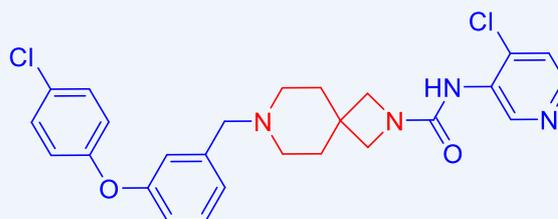
## PharmaBlock Products



PB08259

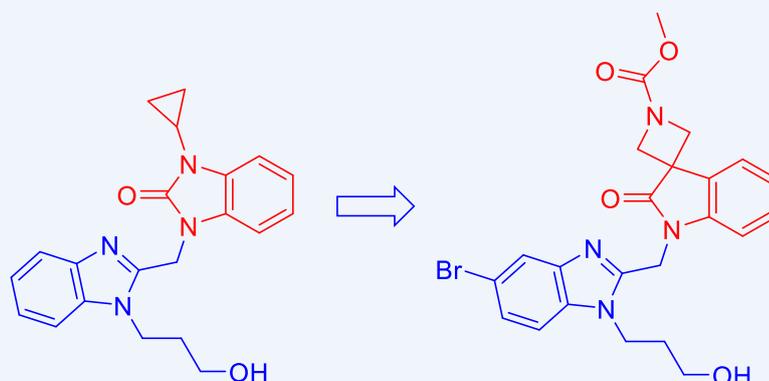


PBLJ0709-1



2,7-diazaspiro[3,5]nonane **19**  
hFAHH IC<sub>50</sub> = 8 nM

Respiratory syncytial virus (RSV) is a major cause of pneumonia and bronchiolitis in young children, immunocompromised adults, and elderly. Unfortunately, there is no effective treatment except an expensive monoclonal antibody (palivizumab) with a dubious safety profile. BMS's pyridinoimidazolone **20** is an efficacious RSV F fusion glycoprotein inhibitor. Using **20** as a starting point, a “patent-busting” or “scaffold-hopping” exercise led to the discovery of a series of novel spiroazetidines 2-oxo-indoline derivatives. The lead compound **21** exhibited excellent *in vitro* potency with an EC<sub>50</sub> value of 0.8 nM and demonstrated 71% oral bioavailability in mice.<sup>15</sup>



pyridinoimidazolone **20**  
RSV EC<sub>50</sub> = 22 nM  
CC<sub>50</sub> >100 μM

spiroazetidine **21**  
RSV EC<sub>50</sub> = 0.8 nM  
CC<sub>50</sub> >100 μM  
%F = 71%

## PharmaBlock Products



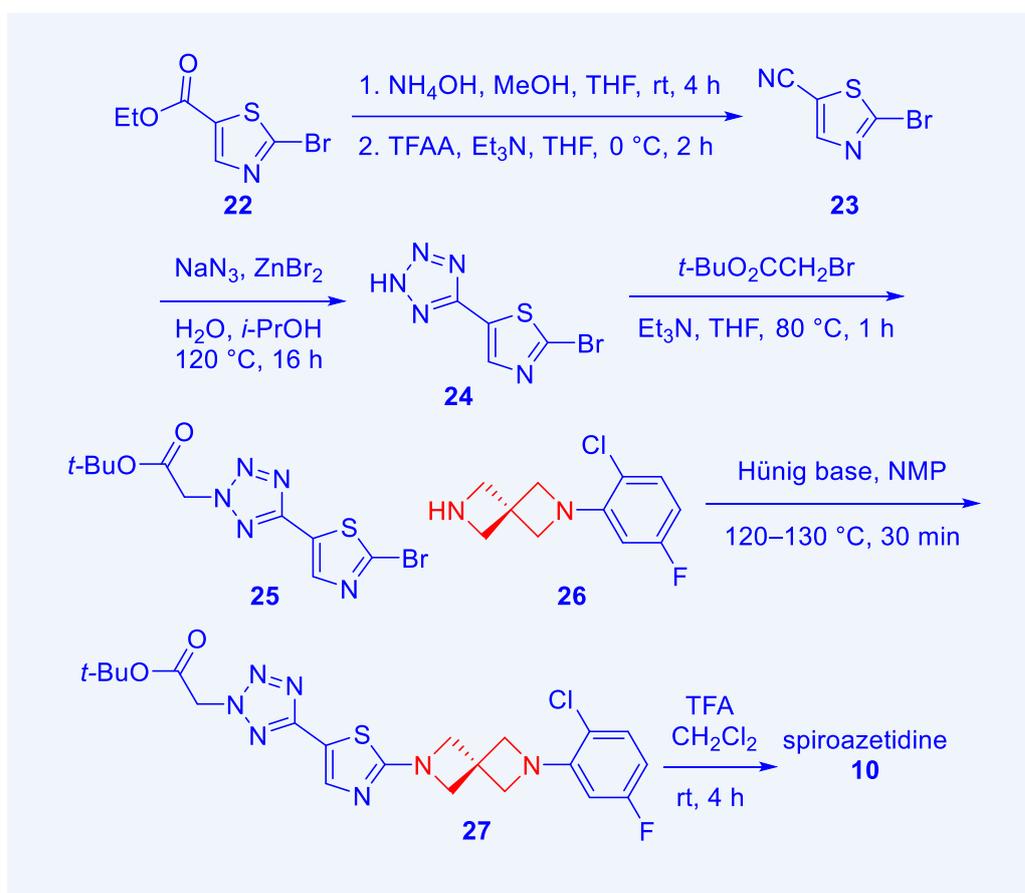
PB05200



PBZ2900

## Synthesis of Some Spiroazetidines

Synthesis of Merck Frost's liver-selective SCD inhibitor spiroazetidines **10** commenced with preparation of bromothiazole-nitrile **23** from bromothiazole-ester **22** in two steps. Tetrazole **24** was then elaborated by condensation of bromothiazole-nitrile **23** with sodium azide.  $S_N2$  alkylation of tetrazole **24** with *t*-butyl bromoacetate gave rise to ester **25**, which was followed by an  $S_NAr$  reaction with aryl-2,6-diazaspiro[3.3]heptane **26** to assemble the adduct **27**. TFA-promoted deprotection of the ester then delivered spiroazetidines **10**.<sup>8</sup>

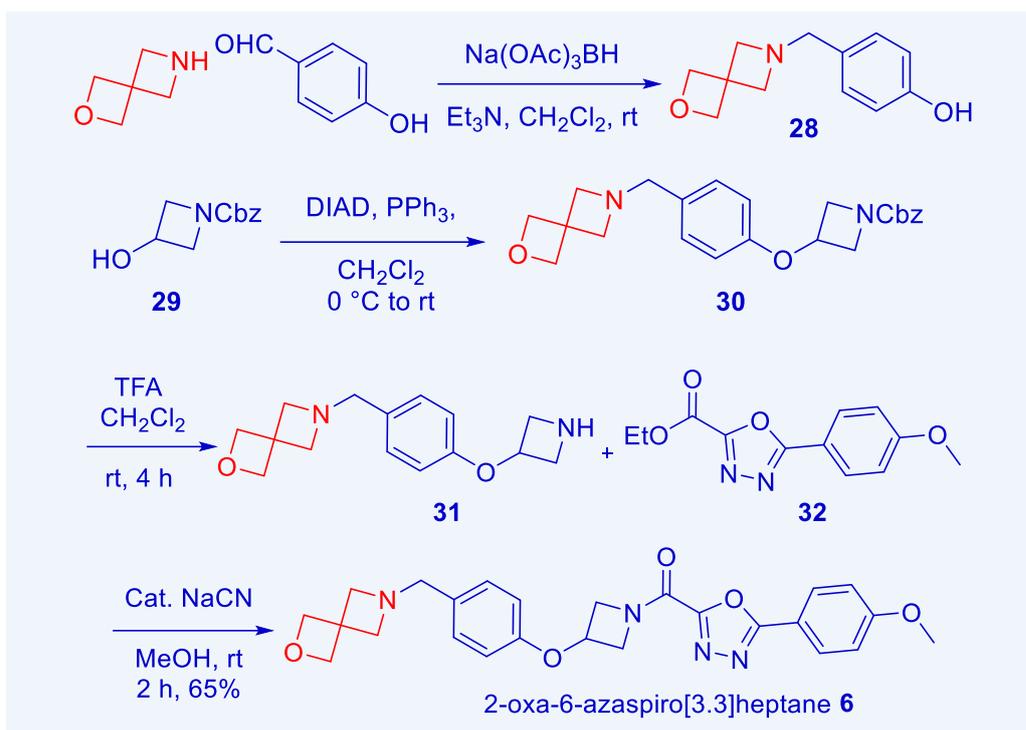


AstraZeneca began synthesis of their MCHR1 antagonist clinical candidate AZD1979 (**4**) with a reductive amination of *p*-anisaldehyde with 2-oxa-6-azaspiro[3.3]heptane to prepare benzylamine **28**. Subsequently, a Mitsunobu reaction between benzylamine **28** and Cbz-protected azetidine **29** provided azetidine ether **30**. After deprotection, the resulting “naked” azetidine **31** was coupled with oxadiazole **32** with the aid of catalytic amount of sodium cyanide to deliver the desired 2-oxa-6-azaspiro[3.3]heptane **4**.<sup>4</sup>



PharmaBlock is recognized for its outstanding capability in the design, synthesis, production and commercialization of novel building blocks for use throughout the drug R&D process.

- 80,000+ building blocks
- 16,000+ in stock in both USA and China
- 20,000+ supplied within two weeks
- 1,000+ SAR tool kits
- Novel building blocks designed upon daily monitoring on recent researches and patents
- Keep optimizing cost effective route for better price and sustainable supply
- Fast delivery of custom synthesis
- Enabling technologies of flow chemistry, biocatalysis, photochemistry, electrochemistry, and fluorination, etc.
- Commercial production with GMP compliance



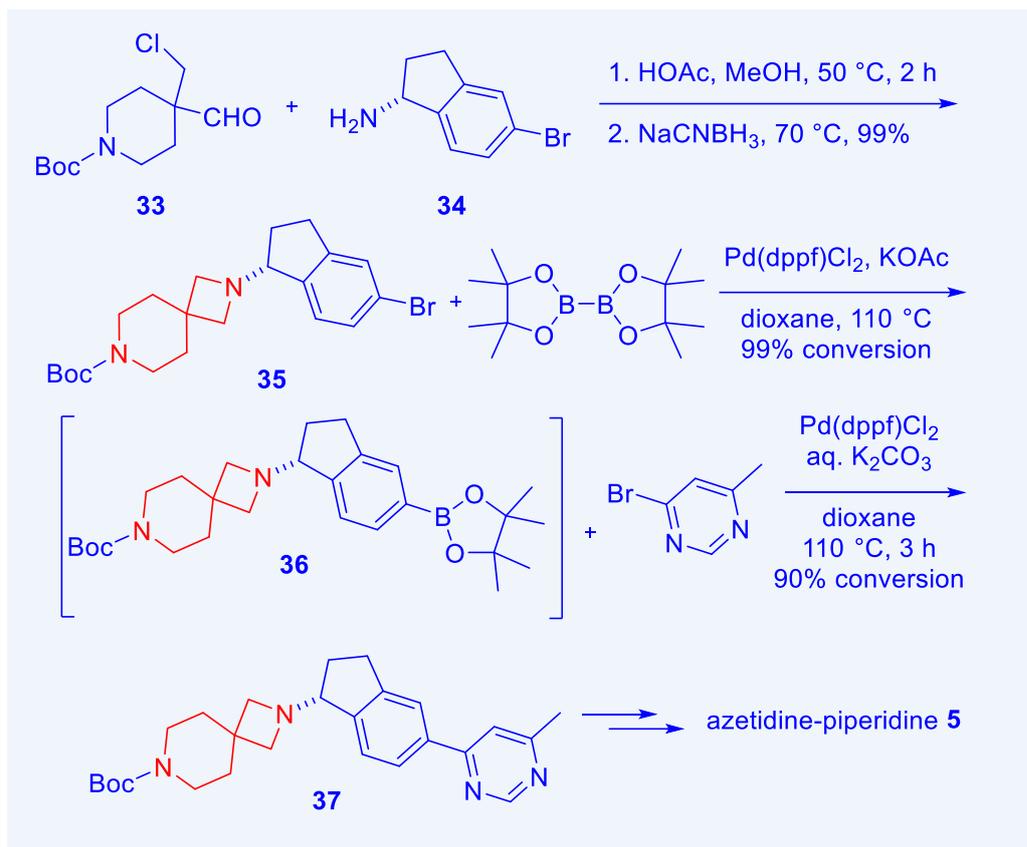
To produce GR inverse agonist PF-5190457 (**5**), Pfizer first carried out a reductive amination of chloroaldehyde **33** with indane-amine **34** to assemble azetidine–piperidine **35**. Scaffold **35** served as a linchpin-like modular framework for further functionalizations. Palladium-catalyzed Miyaura reaction between bromide **35** and bis(pinacolato)diboron produced boronate intermediate **36**, which underwent a Suzuki coupling with 4-bromo-6-methylpyrimidine to provide adduct **37**. After removal of the Boc protective group on **37**, the revealed azetidine–piperidine was coupled with the requisite acid to deliver PF-5190457 (**5**).<sup>5</sup>



### Contact Us

PharmaBlock Sciences  
(Nanjing), Inc.  
Tel: +86-400 025 5188  
Email:  
sales@pharmablock.com

PharmaBlock (USA), Inc.  
Tel (PA): 1-877 878 5226  
Tel (CA): 1-267 649 7271  
Email:  
salesusa@pharmablock.com



For spiroazetidine building blocks that are not commercially available, Carreira<sup>16,17</sup> and Mykhailik<sup>18,19</sup> have published excellent works on their preparations.

To conclude, spiroazetidines, like all spirocyclic scaffolds, are bestowed with three advantages over their flat counterparts: inherent three-dimensional structures may offer more interactions with target proteins; spiroazetidines may provide superior physicochemical properties thus more drug-like; and novel structures may offer fresh intellectual properties. Meanwhile, although no spiroazetidine-containing drugs are current approved for marketing, at least two of them have been advanced to clinical trials. Their applications in medicinal chemistry are destined to grow, especially since many of them are now commercially available.

## References

1. (a) Lovering, F.; Bikker, J.; Humblet, C. *J. Med. Chem.* **2009**, *52*, 6752–6756. (b) Lovering, F. *MedChemComm* **2013**, *4*, 515–519.
2. (a) Zheng, Y. J.; Tice, C. M. *Exp. Opin. Drug Discov.* **2016**, *11*, 831–834. (b) Zheng, Y. J.; Tice, C. M.; Singh, S. B. *Bioorg. Med. Chem. Lett.* **2014**, *24*, 3673–3682.
3. Dhillon, S. *Drugs* **2020**, *80*, in press.
4. Johansson, A.; Löfberg, C.; Antonsson, M.; von Unge, S.; Hayes, M. A.; Judkins, R.; Ploj, K. arolina; Benthem, L.; Linden, D.; Brodin, P.; et al. *J. Med. Chem.* **2016**, *59*, 2497–2511.
5. Bhattacharya, S. K.; Andrews, K.; Beveridge, R.; Cameron, K. O.; Chen, C.; Dunn, M.; Fernando, D.; Gao, H.; Hepworth, D.; V. Jackson, M.; Khot, V.; et al. *ACS Med. Chem. Lett.* **2014**, *5*, 474–479.
6. Krivtsov, A. V.; Evans, K.; Gadrey, J. Y.; Eschle, B. K.; Hatton, C.; Uckelmann, H. J.; Ross, K. N.; Perner, F.; Olsen, S. N.; Pritchard, T.; et al. *Cancer Cell* **2019**, *536*, 660–673.
7. Maryanoff, B. E.; O'Neill, J. C.; McComsey, D. F.; Yabut, S. C.; Luci, D. K.; Jordan, A. D.; Masucci, J. A.; Jones, W. J.; Abad, M. C.; Gibbs, A. C.; et al. *ACS Med. Chem. Lett.* **2011**, *2*, 538–543.
8. Lachance, N.; Gareau, Y.; Guiral, S.; Huang, Z.; Isabel, E.; Leclerc, J.-P.; Leger, S.; Martins, E.; Nadeau, C.; Oballa, R. M.; et al. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 980–984.
9. Kwong, A. D.; Kauffman, R. S.; Hurter, P.; Mueller, P. *Nat. Biotechnol.* **2011**, *29*, 993–1003.
10. Bondada, L.; Rondla, R.; Pradere, U.; Liu, P.; Li, C.; Bobeck, D.; McBrayer, T.; Tharnish, P.; Courcambeck, J.; Halfon, P.; et al. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 6325–6330.
11. Henderson, J. L.; Kormos, B. L.; Hayward, M. M.; Coffman, K. J.; Jasti, J.; Kurumbail, R. G.; Wager, T. T.; Verhoest, P. R.; Noell, G. S.; Chen, Y.; et al. *J. Med. Chem.* **2015**, *58*, 419–432.
12. Kung, D. W.; Coffey, S. B.; Jones, R. M.; Cabral, S.; Jiao, W.; Fichtner, M.; Carpino, P. A.; Rose, C. R.; Hank, R. F.; et al. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 4281–4287.
13. McClure, K. F.; Jackson, M.; Cameron, K. O.; Kung, D. W.; Perry, D. A.; Orr, S. T.; Zhang, Y.; Kohrt, J.; Tu, M.; Gao, H.; et al. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 5410–5414.
14. Keith, J. M.; Jones, W. M.; Pierce, J. M.; Seierstad, M.; Palmer, J. A.; Webb, M.; Karbarz, M. J.; Scott, B. P.; Wilson, S. J.; Wennerholm, M. L.; et al. *Bioorg. Med. Chem. Lett.* **2014**, *24*, 737–741.
15. Shi, W.; Jiang, Z.; He, H.; Xiao, F.; Lin, F.; Sun, Y.; Hou, L.; Shen, L.; Han, L.; Zeng, M.; et al. *ACS Med. Chem. Lett.* **2018**, *9*, 94–97.
16. Guérot, C.; Tchitchanov, B. H. T.; Knust, H.; Carreira, E. M. *Org. Lett.* **2011**, *11*, 780–783.
17. Carreira, E. M.; Fessard, T. C. *Chem. Rev.* **2014**, *114*, 8257–8322.
18. Kirichok, A. A.; Shton, I.; Kliachyna, M.; Pishel, I.; Mykhaiiuk, P. K. *Angew. Chem. Int. Ed.* **2017**, *56*, 8865–8869.
19. Kirichok, A. A.; Shton, I.; Pishel, I.; Zozulya, S. A.; Borysko, P. O.; Kubyshkin, V.; Zaporozhets, O. A.; Tolmachev, A. A.; Mykhaiiuk, P. K. *Chem. Eur. J.* **2018**, *24*, 5444–5449.